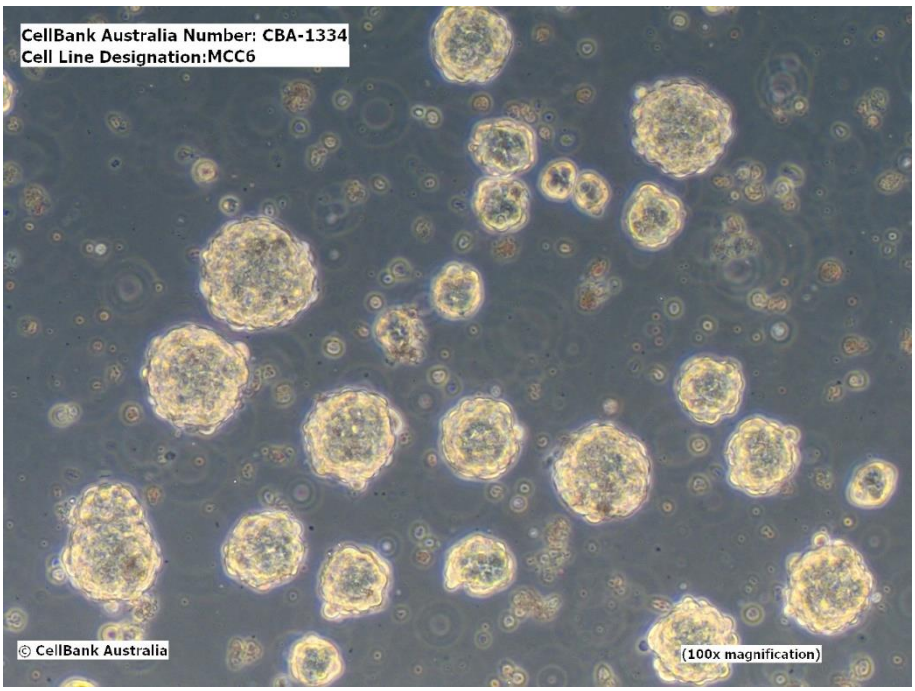
Confidence in cell lines. Integrity in research.

Cell Line Information Sheet – MCC6

CellBank Catalogue No.:	**CBA-1334**
Lot Number:	**13341219E**
Passage Number:	51
Total Cell Number:	Approximately 5 x 10^6 cells
Cell Line Description:	Human Merkel cell carcinoma cell line, derived from a secondary tumour of the lymph node of a 70yr old male patient
Organism:	Human (*Homo sapiens*)
Tissue:	Metastatic, derived from lymph node.
Growth Properties:	Suspension culture, slow growing. Doubling time is more than 1 week.
Morphology:	Small cells in tight clusters and balls
Image:	
Growth Medium:	RPMI 1640 (with 2mM L-Glutamine + 25mM HEPES) + 15% Foetal Bovine Serum.+10 ug/ml Insulin, 5.5 ug/ml Transferrin, 5 ng/ml Sodium Selenite (ITS) (ITS :Sigma Catalogue number I3146-5ML 100x solution.)

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Resuscitation	**Remove protective cryoflex layer around the ampoule prior to thawing.** Thaw the ampoule by gently agitating in a 37ºC waterbath; thawing should be rapid (around 2 minutes). A centrifugation step to remove the cryoprotectant after thawing is necessary for this cell line. Seed 1 vial of **MCC6 Lot #13341219E into 1xT25 flask,** after 3-4 days it can be transferred to a T75 flask with the addition of 50% fresh medium. Note that this cell line can be challenging to resuscitate from thaw, is slow growing and favours conditioned medium.
Subculturing Procedure:	**Medium Renewal:** 1-2 times per week. Feed culture by dilution of 50% fresh medium with 50% conditioned medium where possible.This cell line grows optimally with some conditioned medium **Subcultivation ratio**: 1:2, Seed 1 vial of **MCC6 Lot #13341219E into 1xT25 flask** Split cultures when they reach high density by dilution 1:2. Break up the clusters to smaller clumps by pipetting up and down several times. Do not break the culture down to single cells. **Culture conditions:** Incubate the culture at 37ºC with 5% CO_2. **Cryoprotectant Medium:** 10% DMSO + 90% FCS
Safety Precaution:	Where cell lines are shipped as frozen ampoules, there is a small risk that the ampoule may be pressurised due to the expansion of trapped liquid nitrogen and could explode on warming. It is recommended that persons handling ampoules of frozen cells wear appropriate personal protective equipment including laboratory coat, insulated gloves and a full protective face shield.
Handling Procedure for Frozen Cells:	Upon receipt, frozen ampoules should be transferred directly to liquid nitrogen storage without delay, if not to be used immediately. Storage at -80°C may result in loss of viability.
Biosafety Level:	Cell line of human origin. CellBank Australia recommends that cell lines be handled at category PC-2* containment level. *AS/NZS 2243.3:2010

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Additional Information:	Does not grow well when seeded at a low density or when clusters are broken down to single cells. This cell line was originally derived and cultured using a feeder cell line. CellBank Australia has cultured the cell line without the use of a feeder layer but with the addition of insulin, transferrin and selenite (ITS) to the base medium of RPMI1640 and 10%FBS.
Depositor:	Helen Leonard – Queensland Institute of Medical Research, Australia.
References:	**Original Reference** **Leonard, J.H.**, Bell, J.R. and Kearsley, J.H., Characterization of cell lines established from Merkel cell (small cell) cancer of the skin. Int. J. Cancer, 55, 803-810, 1993. PMID: 8244578
Use Restrictions:	These cells are distributed for research purposes only - refer to the Sales Terms and Conditions.
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